

***In vitro* and *in vivo* properties of distinct populations of amniotic fluid mesenchymal progenitor cells**

**Maria G. Roubelakis^{a, b, *, #}, Vasiliki Bitsika^{a, b, #}, Dimitra Zagoura^{a, b}, Ourania Trohatou^{a, b},
Kalliopi I. Pappa^{b, c}, Manousos Makridakis^d, Aristidis Antsaklis^c,
Antonia Vlahou^d, Nicholas P. Anagnou^{a, b, *}**

^aCell and Gene Therapy Laboratory, Centre of Basic Research, Biomedical Research Foundation,
Academy of Athens (BRFAA), Athens, Greece

^bLaboratory of Biology, University of Athens School of Medicine, Athens, Greece

^cFirst Department of Obstetrics and Gynecology, University of Athens School of Medicine, Athens, Greece

^dBiotechnology Laboratory, Centre of Basic Research, Biomedical Research Foundation,
Academy of Athens (BRFAA), Athens, Greece

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Abstract

Human mesenchymal progenitor cells (MPCs) are considered to be of great promise for use in tissue repair and regenerative medicine. MPCs represent multipotent adherent cells, able to give rise to multiple mesenchymal lineages such as osteoblasts, adipocytes or chondrocytes. Recently, we identified and characterized human second trimester amniotic fluid (AF) as a novel source of MPCs. Herein, we found that early colonies of AF-MPCs consisted of two morphologically distinct adherent cell types, termed as spindle-shaped (SS) and round-shaped (RS). A detailed analysis of these two populations showed that SS-AF-MPCs expressed CD90 antigen in a higher level and exhibited a greater proliferation and differentiation potential. To characterize better, the molecular identity of these two populations, we have generated a comparative proteomic map of SS-AF-MPCs and RS-AF-MPCs, identifying 25 differentially expressed proteins and 10 proteins uniquely expressed in RS-AF-MPCs. Furthermore, SS-AF-MPCs exhibited significantly higher migration ability on extracellular matrices, such as fibronectin and laminin *in vitro*, compared to RS-AF-MPCs and thus we further evaluated SS-AF-MPCs for potential use as therapeutic tools *in vivo*. Therefore, we tested whether GFP-lentiviral transduced SS-AF-MPCs retained their stem cell identity, proliferation and differentiation potential. GFP-SS-AF-MPCs were then successfully delivered into immunosuppressed mice, distributed in different tissues and survived longterm *in vivo*. In summary, these results demonstrated that AF-MPCs consisted of at least two different MPC populations. In addition, SS-AF-MPCs, isolated based on their colony morphology and CD90 expression, represented the only MPC population that can be expanded easily in culture and used as an efficient tool for future *in vivo* therapeutic applications.

Keywords: amniotic fluid • MPCs • migration • proteomic analysis • *in vivo* properties

[#]Joint first authors.

*Correspondence to: Nicholas P. ANAGNOU, M.D., Ph.D.,
Professor of Biology and Head of Laboratory of Biology
University of Athens School of Medicine, Athens 115 27, Greece and
Group Leader, Laboratory of Cell and Gene Therapy
Foundation for Biomedical Research of the Academy of Athens (IIBEAA),
Athens 115 27, Greece
Tel.: +30-210-746-2341; +30-210-746-2356; +30-210-6597-013
Fax: +30-210-746-2412
E-mail: anagnou@med.uoa.gr
<http://www.bioacademy.gr/lab/lab.php?lb=36>

and
Maria G. ROUBELAKIS, Ph.D.
University of Athens School of Medicine,
Athens 115 27, Greece and
Foundation for Biomedical Research
of the Academy of Athens (IIBEAA),
Athens 115 27, Greece
Tel.: +30-210-6597-013
Fax: +30-210-6597-545
E-mail: mrroubelaki@bioacademy.gr
<http://www.bioacademy.gr/lab/lab.php?lb=36>

Introduction

Adult bone marrow (BM) mesenchymal progenitors cells (MPCs) or mesenchymal stem cells (MSCs), initially described as precursors of fibroblasts or stromal cells, can be isolated taking advantage of their adhesive properties and can be further expanded in culture. Previous studies demonstrated that MPC populations derived from BM are heterogeneous and contain at least two morphologically distinct subpopulations of cells: (a) spindle-shaped (SS), rapidly self-renewing MPCs and (b) flattened-shaped, slowly self-renewing MPCs [1–4]. More interestingly, this subset of SS MPCs is able to preferentially engraft in mice; thus, they appear more promising tools for clinical applications [5]. Similarly, SS and flattened-shaped MPCs were also isolated from umbilical cord blood (UCB) at clonal level [6], with SS subpopulation exhibiting high expression levels of CD90, whereas the flattened was negative for the same antigen [6].

Recently, our group and others [7–9] have isolated MPCs from an alternative source, the second trimester amniotic fluid (AF), which can be obtained during routine amniocentesis without any ethical concerns [7, 10–12]. We characterized these cells based on their phenotype, multipotency, differentiation potential and on their proteomic profile, constructing a two-dimensional electrophoresis (2-DE) proteomic database of AF-MPCs [7]. Most importantly, AF-MPCs were easily isolated and grew more rapidly under the appropriate culture conditions compared to BM-MPCs [7]. In addition, concurrent studies showed that AF-MPCs, seeded in a scaffold and exposed to osteogenic-inducing medium, were able to form bone after subcutaneous implantation *in vivo*, demonstrating an important role in pre-clinical studies [9].

However, AF-MPCs represent a heterogeneous population composed of multiple categories of adherent cells based on morphological, biochemical and growth characteristics [13–16]. Until now, there is no surface epitope or proposed protocol that can distinguish MPCs *in vitro* and *in vivo* [2]. Therefore, most experiments have been carried out with heterogeneous populations of AF-MPCs [7, 8, 11, 12, 16]. Questions regarding the heterogeneity, the mobilization and homing properties of these cells *in vitro* and *in vivo*, as well as their use as carriers of therapeutic agents are still undetermined.

Therefore, a large number of parameters must be taken under consideration in isolating the rapidly expanding MPCs from AF. First, an important determinant of successful stem cell therapeutic transplantation is the ability of the transplanted cells to home, migrate, efficiently engraft and repair damaged tissues. Various molecules have been shown to regulate these processes in MPCs. Among these are the VLA-5 integrin, which comprises of $\alpha 5$ (CD49e) and $\beta 1$ (CD29) subunits and plays key role in migration [7, 8, 11, 17] and the adhesion molecule CD44, which is highly expressed in most of the mesenchymal cell types [18, 19].

Another parameter involves the feasibility of employing genetic modifications on AF-MPCs by introducing therapeutic genes of

interest. For these manipulations, apart from adenoviruses, that have been previously utilized in AF-MPCs [20], lentiviral vectors represent a highly efficient and long-term infection system for transducing stem cells, without the presence of silencing effects during multiple divisions [21–23].

Herein, we have extended our previous findings [7], by isolating and characterizing two different types of AF-MPCs at passage 0 based on their shape, antigen expression levels and functional properties. We named these cells according to their morphology as SS and round-shaped (RS) AF-MPCs. SS-AF-MPCs are characterized by a greater proliferation and multilineage differentiation potential, while they are exhibiting higher levels of CD90 compared to RS-AF-MPCs. Therefore, in the current experiments, we compared their molecular identity, the differentiation capacity and the *in vitro* adhesion properties of both subpopulations. We further analysed the migratory ability, the efficient gene modification and the perspective utilization of SS-AF-MPCs in pre-clinical studies *in vivo*.

Material and methods

Isolation and culture of MPCs from human AF

Human AF-MPCs were isolated from 95 AF samples, collected during scheduled amniocentesis between the 15th and 18th week of gestation [7]. Briefly, second trimester AF samples were obtained, following a written informed consent, approved by the Ethical Committee of the Alexandra Hospital, Athens and the Bioethics Committee of the University of Athens School of Medicine, during scheduled amniocenteses between the 15th and 18th week of gestation. Amniocentesis was performed under aseptic conditions. Using a 22G needle and under ultrasonographic control, 10–15 ml of AF was aspirated for each sample. The procedure-related spontaneous abortion rate ranges from 0.06% to 0.5% [24, 25]. Samples were collected and 10 ml of each were centrifuged at 1300 rpm for 10 min. The pellet was resuspended in Dulbecco's modified Eagle's medium (DMEM; Sigma-Aldrich, St Louis, MO, USA) supplemented with 20% (v/v) foetal bovine serum (FBS) (Gibco-BRL, Paisley, Scotland, UK), in a 25 cm² tissue culture-treated flask and incubated at 37°C in a 5% humidified CO₂ chamber for approximately 20 days, where the first colonies appeared. The medium was then changed every 5 days. The cells were expanded into higher passages and frozen until use.

Colony forming unit-fibroblast (CFU-F) assay

The CFU-F assay was performed by plating 1×10^5 AF cells/well into six-well plates from six randomly selected samples, in the presence of DMEM (Sigma-Aldrich) supplemented with 20% (v/v) FBS (Gibco-BRL). After 18–20 days of culture, CFUs were formed, mechanically selected and sub-cultured separately [26]. The cells from each colony were expanded into higher passages and frozen until use.

Karyotyping

For karyotyping, RS-AF-MPCs and SS-AF-MPCs from four different samples each, grown in log phase, were harvested and karyotyped using Giemsa stain GTG banding, at passage 5. For SS-AF-MPCs, karyotype was examined at passage 32 as well. Forty metaphase spreads were fully analysed and karyotyped in each case.

Antibodies and flow cytometry analysis (FACS)

RS-AF-MPCs and SS-AF-MPCs of passage 3–17 were characterized by FACS analysis using CD90, CD166, CD73, CD105, CD44, c-kit, CD34, CD133, CD31, CD29, CD49e, CD45, CD50, CD106, CD11a, CD62P, CD1a, CD13, CD14, CD62E and CD146 [Becton Dickinson (BD), San Jose, CA, USA], SSEA-4 (Abcam, Cambridge, UK), HLA-ABC and HLA-DR (BD) mouse anti-human monoclonal antibodies or appropriate isotype controls (BD). The latter reactions were developed with fluorescein isothiocyanate (FITC)-conjugated goat anti-mouse IgG secondary antibody (DAKO DakoCytomation Ltd., Cambridgeshire, UK). For apoptosis detection, cells were stained with Annexin V-PE (BD) according to manufacturer's instructions. Cells were analysed using a Beckman Coulter Cytomics FC 500 flow cytometer (Beckman Coulter Ltd., Palo Alto, CA, USA). For immunofluorescent staining, anti-collagen type I (Sigma-Aldrich), anti-E-cadherin (Abcam), anti-N-cadherin (Santa Cruz Biotechnology Inc. Santa Cruz, CA, USA) and anti-Vimentin (Thermo Scientific, Fremont, CA, USA) antibodies were used in fixed cells with 4% paraformaldehyde (Sigma-Aldrich). The latter reactions were developed with FITC-conjugated goat anti-mouse IgG (DAKO DakoCytomation Ltd.) or Alexa-488-conjugated goat anti-rabbit (Invitrogen Ltd., Paisley, UK) secondary antibodies, where appropriate. Isotype-matched antibodies were used as negative control. For Oct-4 and Sox-2 nuclear immunofluorescent staining, cells were permeabilized in 0.2% (v/v) Triton-X (Sigma-Aldrich) for 30 min at room temperature after fixation as described above. Cells were then stained with rabbit anti-human Oct-3/4 (Santa Cruz Biotechnology Inc.) or rabbit anti-human Sox-2 (Millipore, CA, USA) antibodies. The latter reactions were developed with Alexa-488-conjugated goat anti-rabbit (Invitrogen Ltd.) secondary antibody. Slides were mounted with Vectashield mounting medium (Vector Laboratories Inc., CA, USA) containing DAPI solution. Cells were visualized and photographed on a Leica CTR-MIC Fluorescent microscope.

MTS proliferation assay

MTS assay was performed to estimate the proliferation rate of SS-AF-MPCs, RS-AF-MPCs or GFP-SS-AF-MPCs. Cells were plated at a density of 10^3 per well in a 96-well plate and cultured for 6–7 days in the presence of DMEM (20% FBS) in five replicates. Appropriate amount of MTS (Promega Ltd, Madison, WI, USA) was added to each well and incubated for 3 hrs. The absorbance was recorded at 490 nm using an ELISA plate reader (ELX 800; Biotek Instruments Inc, VT, USA). The percentage increase of proliferation was calculated using the following formula: $[(OD_{dayx} - OD_{day0}) / OD_{day0} \times 100]$. Statistical analysis was performed using the Student's t-test. Three individual samples from each cell type were analysed.

For conditioned medium (CM) preparation, SS-AF-MPCs were cultured until 80% confluency and further allowed to grow in a medium containing 20% FBS for 48 hrs. CM was collected, filtered and stored at -20°C until use. RS-AF-MPCs were cultured in CM derived from SS-AF-MPCs for 6 days. CM was changed every day.

For temperature proliferation analysis, SS-AF-MPCs at passage 15–20 were cultured for 1 and 6 days in DMEM (20% FBS) at 37°C and 33°C , respectively.

Transwell migration assay

SS-AF-MPCs (passage 5–15) and RS-AF-MPCs (passage 2–5) were cultured for 48 hrs in DMEM (2% FBS) and then were transferred to the insert of a 8 mm pore size transwell plate, at $5 \times 10^4 / 100 \mu\text{l}$ density (Corning-Costar, Cambridge, MA, USA). The cells were then allowed to migrate for 16 hrs across the pore membrane, towards human plasma fibronectin (20 $\mu\text{g}/\text{ml}$; Sigma-Aldrich), laminin I (20 $\mu\text{g}/\text{ml}$; Sigma-Aldrich) or DMEM (2% FBS). The non-migrated cells were removed from the top of the insert with a cotton swab. The migrated cells were then fixed with 4% paraformaldehyde (Sigma-Aldrich) on the membrane and stained using the Ral Kit (Ral Reactif, Paris, France). For migration blocking experiment, cells were pre-incubated with CD44 (clone 515, BD). Migration was quantified by counting the stained nuclei that passed through the filter. Photographs were taken from a minimum of 10 fields of view ($20\times$) for each membrane and then counted by using Image J software. Three independent experiments were performed each including three replicates. Statistical analysis was performed using the Student's t-test.

Adhesion assay

SS-AF-MPCs (passage 5–15) and RS-AF-MPCs (passage 2–5) were pre-incubated either with CD44 (clone 515, BD), CD49e (blocking antibody, BD) or IgG1 isotype control (BD) and then transferred to a 24-well plate, coated with fibronectin (Sigma-Aldrich) or hyaluronic acid (Sigma-Aldrich). Non-treated SS- and RS-AF-MPCs were used as positive controls. Cells were let to adhere for 2 hrs and then the supernatant was removed. The adherent cells to the bottom of the well were fixed with 4% (w/v) paraformaldehyde (Sigma-Aldrich), stained using the Ral Kit (Ral Reactif). Photographs were taken from a minimum of 10 fields of view ($20\times$) for each membrane and then counted by using Image J software. Three independent experiments were performed, each including three replicates. Statistical analysis was performed using the Student's t-test.

RT-PCR procedure

Total RNA was extracted from the cells using the Tri Reagent (Sigma-Aldrich) according to the manufacturer's instructions. The mRNA was reverse transcribed to cDNA using the M-MLV Reverse Transcriptase-RNase H Minus kit (Promega Ltd.).

Real-time quantitative PCR and semi-quantitative PCR

Real-time quantitative PCR was conducted on an ABI Prism 7700 apparatus [Applied Biosystems (ABI), Foster City, CA, USA]. Each cDNA sample was mixed with specific primer sets and PCR master mix (Applied Biosystems, No. 4312704). TaqMan RT-PCR was used for oct-4, nanog and sox-2 gene expression analysis with primers and conditions designated by Assays on DemandTM, Gene Expression Products [ABI Hs00742896_s1 (oct-4),

Hs02387400_s1 (nanog), Hs01053049_s1 9 (sox-2), respectively]. Data were analysed with the ABI Prism 7700 SDS software (ABI). The expression of oct-4, nanog and sox-2 was normalized to GAPDH internal control (GAPDH, ABI, No. 433764T), as described by Zannettino *et al.* [27]. The levels of gene expression were normalized after subtracting the Ct value of the GAPDH internal control from that of the x (x = oct-4, nanog or sox-2) Ct value for SS-AF-MPCs and RS-AF-MPCs ($\Delta Ct = |Ct_x (SS-AF-MPCs \text{ or } RS-AF-MPCs) - Ct_{GAPDH}|$). To compare the levels of oct-4, nanog and sox-2 expression between SS-AF-MPCs and RS-AF-MPCs, the $\Delta\Delta Ct$ value was determined using the formula ($\Delta\Delta Ct = \Delta Ct_x(SS-AF-MPCs) - \Delta Ct_x(RS-AF-MPCs)$). Then, the relative level of oct-4, nanog and sox-2 in SS-AF-MPCs was compared to RS-AF-MPCs by setting the oct-4, nanog and sox-2 expression in RS-AF-MPCs value to 1 and determining the fold change in expression against this value using the following formula: $2^{\Delta\Delta Ct}$.

PCR reaction for albumin was performed using cDNA, primer pairs and Go Taq[®] Green master mix (Promega Ltd.). The primers used were: F: 5'-AGATGACAACCCAAACCTCCC-3' and R: 5'-CAGCAGCACGACAGTAATC-3'. The semi-quantitative PCR analysis was determined by using the Image J software after normalization to the β -actin endogenous control (F: 5'-TCTACAATGAGCTGCGTGTG-3' and R: 5'-CAACTAAGTCATAGCCGCC-3').

Differentiation assays

Osteogenic differentiation

To induce osteogenic differentiation SS-AF-MPCs or RS-AF-MPCs of passage 5–7 were cultured to 60–70% confluency in osteogenic medium for 3 weeks, as described previously [7]. The differentiation potential for osteogenesis was assessed by alkaline phosphatase staining using BCIP/NBT reagent (Sigma-Aldrich), according to the manufacturer's instructions.

Adipogenic differentiation

SS-AF-MPCs or RS-AF-MPCs of passage 5–13, grown as described above, were treated with adipogenic medium for 3 weeks, as described previously [7]. Medium changes were carried out twice weekly. For the determination of adipogenic differentiation, formation of intracellular lipid droplets was monitored under microscope and was confirmed by Oil Red O staining (Sigma-Aldrich).

Chondrogenic differentiation

To induce chondrogenic differentiation, SS-AF-MPCs or RS-AF-MPCs of passage 5–10, were cultured in high-density pellet mass cultures. Briefly, 2.5×10^5 cells were placed into a conical polypropylene tube with 0.5 ml of defined medium, consisting of DMEM (Sigma-Aldrich) supplemented with 5.33 μ g/ml linoleic acid (Sigma-Aldrich), insulintransferrin-selenium⁺ (ITS⁺) (Sigma-Aldrich), 1.25 mg/ml bovine serum albumin (Sigma-Aldrich), 10 ng/ml transforming growth factor β 3 (TGF- β 3) (Peprotech), 10^{-7} M dexamethasone (Sigma-Aldrich), 0.17 mM ascorbate acid (Sigma-Aldrich) and 0.35 mM L-Proline (Sigma-Aldrich). Cells were centrifuged at 2000 rpm for 5 min and maintained at 37°C for 21 days. Medium was changed twice a week. After 21 days of pellet mass culture, the pellets were fixed in 10% formalin (Sigma-Aldrich), paraffin embedded, sectioned and stained with Alcian Blue (Sigma-Aldrich) and haematoxylin and eosin (Sigma-Aldrich).

Hepatogenic differentiation

For hepatogenic differentiation, SS-AF-MPCs or RS-AF-MPCs of passage 5–13, at $1.0\text{--}1.3 \times 10^4/\text{cm}^2$ density, were treated with appropriate hepatogenic medium, as described previously [7]. Hepatogenesis was assessed

after 3 weeks by periodic acid-Schiff (PAS) staining (Sigma-Aldrich), measurement of urea concentration (Urea UV Liquid, Technologia Diagnostics, Greece) using the Chemwell 2910 autoanalyzer (Awareness Technology Inc., Palm City, FL, USA) and determination of albumin expression by semi-quantitative PCR.

Osteogenic, adipogenic and PAS hepatogenic differentiation assays were quantified by microscopic analyses of 10 fields per image using the Image J software.

Proteomic analysis–mass spectrometry (MS)

Three samples of SS- or RS-AF-MPCs of passage 5–10 were analysed by 2D-gel electrophoresis in duplicate each, as described before [7]. Protein spots were manually excised, tryptic digested and Peptide Mass Fingerprinted [7]. Stringent criteria were used for protein identification with a maximum allowed mass error of 25 ppm (parts per million) and a minimum of five matching peptides [29]. Notably, a high percentage of the proteins were identified based on 10 matches. The probability of a false identity was usually lower than 10^{-5} . All selected proteins were identified and further analysed for their functional properties by the Swiss-Prot (<http://expasy.org/sprot/>) and Human Protein Reference Databases (<http://www.hprd.org/>). 2DE image analysis was performed by the use of the PD-Quest 8.0 (BioRad, Hercules, CA, USA) software package. Protein expression was shown as a ratio of the intensity of protein spots in SS-AF-MPCs to RS-AF-MPCs or vice versa in ppm. Comparison of the expression level of the various protein spots was conducted by the use of Student's t-test and Mann-Whitney test. In all cases, $P < 0.05$ (95% confidence levels) was considered statistically significant.

Western blot

Total proteins of SS-AF-MPCs and RS-AF-MPCs were separated by 10% SDS-PAGE and electroblotted to Hybond-ECL NC membrane (Amersham Biosciences, Sweden). Protein extracts were derived from a pool of three SS-AF-MPCs or RS-AF-MPCs individual samples of different passages, respectively. After blocking, membranes were incubated overnight at 4°C with the primary antibodies: mouse anti-human CK18 (DakoCytomation), mouse anti-human Cathepsin (BD) or mouse anti-human CK19 (DakoCytomation). Mouse anti-human β -actin antibody (Sigma-Aldrich) was used as a control of equal loading. Membranes were then incubated with anti-mouse HRP-conjugated secondary antibody (Santa Cruz Biotechnology Inc.) and developed by ECL (Perkin-Elmer, MA, USA) detection system. Films were scanned and images were analysed using Quantity One software (BioRad).

Lentiviral vector generation, production and transduction of SS-AF-MPCs

The four plasmid expression lentiviral system containing the pCCLsin.PPT.hPGK.GFP plasmid [28] was kindly gifted by Prof. L. Naldini and used for enhanced GFP expression. Virus was produced by transient transfection into 293T cells, as previously described [29], and collected by ultracentrifugation using an Ultracentrifuge Discovery 100 Sorvall (Thermo Fisher Scientific Inc., Waltham, MA, USA). The concentrated virus was resuspended in PBS supplemented with 0.5% BSA (Sigma-Aldrich). The lentiviral titres were determined by infection of HT1080 cells with serial dilutions of the concentrated viral stock. GFP fluorescent cells were

1 identified by fluorescent microscopy and FACS analysis. Titers ranged from
 2 5×10^8 to 3×10^9 infectious units (IU)/ml. Approximately, 5×10^4 per
 3 well SS-AF-MPCs were seeded in a six-well plate 1 day in advance. Virus
 4 was added in a multiplicity of infection (MOI) of 10–100.

6 ***In vivo* engraftment of GFP-SS-AF-MPCs**

7 NOD-SCID mice were kindly provided by Drs G. Vassilopoulos and E. Siapati
 8 and were housed and maintained at the Animal Facility of the Biomedical
 9 Research Foundation of the Academy of Athens (BRFAA). The procedures for
 10 the care and treatment of animals were performed according to the institu-
 11 tional guidelines, which follow the guidelines of the Association for
 12 Assessment and Accreditation of Laboratory Animal Care (AAALAC) and the
 13 recommendations of the Federation of European Laboratory Animal Science
 14 Associations (FELASA) and approved by the Institutional (BRFAA) Animal
 15 Care and Use Committee. Six to eight weeks old animals ($n = 8$) received
 16 intravenously (i.v.) by tail vein injection 1×10^6 GFP-SS-AF-MPCs (passages
 17 15–40). The animals were sacrificed 4 and 10 days later and the tissues were
 18 analysed by immunohistochemistry, FACS and RT-PCR. For the detection of
 19 GFP transgene, genomic DNA was isolated from each organ/tissue collected,
 20 using Proteinase K (Sigma-Aldrich), followed by phenol–chloroform extrac-
 21 tion. RT-PCR analysis was carried out using primers designed on the WPRE
 22 sequence of the lentiviral vector present only in the infused GFP-SS-AF-MPCs
 23 (F: 5'-T T C T C C T C C T T G T A T A A A T C C T G G T T-3' and R: 5'-C G C
 24 C A C G T T G C C T G A C A-3') and SYBR master mix (Roche Applied
 25 Sciences, Indianapolis, IN, USA), according to manufacturer's protocol.

26 Further, to assess SS-AF-MPCs viability in matrigel *in vivo*, NOD/SCID
 27 mice of the same age ($n = 6$) received subcutaneously 1×10^6 SS-AF-
 28 MPCs in 200 μ l of matrigel (Sigma-Aldrich) into the tail base. As negative
 29 control, 1×10^6 SS-AF-MPCs in 200 μ l of PBS were used. The animals
 30 were sacrificed 1 and 10 days later and the matrigel mass was excised and
 31 photographed. Cells were then disassociated by the use of 2% (w/v) colla-
 32 genase (Sigma-Aldrich) for 2 hrs at 37°C and analysed by FACS.

33 **Immunohistochemistry**

34 At 4 and 10 days after i.v. injection of GFP-SS-AF-MPCs, animals were sac-
 35 rificed and analysed by immunohistochemistry for GFP or CD90 expres-
 36 sion. Tissues were fixed in 10% formalin (Sigma-Aldrich) and embedded
 37 in paraffin. Non-specific binding was blocked using 10% donkey serum in
 38 PBS. Five-micrometre sections were subsequently incubated with eGFP
 39 (Chemicon, Temecula, CA, USA), CD90 (BD) or appropriate isotype control
 40 antibodies. The reaction was developed with biotinylated goat anti-mouse
 41 secondary antibody (DakoCytomation), followed by ABC-complex-HRP
 42 (DakoCytomation) and DAB (Vector Laboratories Inc.). Slides were coun-
 43 terstained in Gill's haematoxylin (Sigma-Aldrich).

44 **Results**

45 **Phenotypic characterization of two different** 46 **AF-MPC populations**

47 As reported previously, BM-MSC and UCB-MSC cultures contained
 48 distinct cell types differing at shape and size [4, 6]. In this study, cell

49 pellets from 95 second trimester AF samples were plated according
 50 to previous protocols [7] and then plastic adherent cells were
 51 isolated. Microscopic analysis led to the identification of two mor-
 52 phological different adherent MPC types in AF, termed as SS (6%,
 53 five established cell lines) and RS MPCs (94%) (Fig. 1A and B). We
 54 initially noticed that the percentage of proliferation increase in SS-AF-
 55 MPCs and RS-AF-MPCs differed during passages 5–7. A more
 detailed analysis revealed that SS-AF-MPCs exhibited high prolifera-
 tive capacity and were passed over 45 passages to date, whereas
 RS-AF-MPCs exhibited a significantly lower proliferative potential
 and reached up to passage 4–7 (Fig. 1C). More importantly, RS-AF-
 MPCs when cultured in CM derived from SS-AF-MPCs exhibited a
 statistically significant increased proliferative potential (Fig. 1C), sug-
 gesting that these cells may require paracrine factors derived from
 the SS-AF-MPCs for expanding. The identification of these two
 different AF populations is of great importance for potential use of
 SS-AF-MPCs in pre-clinical applications and for this reason we
 attempted to investigate their characteristics in more detail.

We first observed that unselected cultures at passage 0–1 con-
 tained a mixture of SS and RS cells [Fig. 1B(i)]. However, at
 two to three passage in 94% of the sample cases of unselected
 cells, the RS cells were more abundant, overtaking the culture
 [Fig. 1B(ii)]. The rest 6% of the samples represented a SS cell
 population [Fig. 1B(iii)] that can be expanded up to 30–50 pas-
 sages to date with normal karyotype [Fig. S1E(i) and (iii)] and high
 proliferation capacity (Fig. 1C). The reason why the SS-AF-MPCs
 were the predominant and the only population at passages 2–3
 onwards in the 6% of the AF samples only, is still undetermined.
 In an attempt to characterize better these two subpopulations, we
 randomly chose six AF samples at passage 0, where a mixture of
 both cell types existed at almost equal frequency. We then
 mechanically isolated 20 individual CFUs, 10 with round and 10
 with SS morphology in total (Fig. 1A). The clonal SS-AF-MPCs and
 RS-AF-MPCs were further expanded *in vitro* and retained their
 morphological characteristics during culture.

The cell surface antigenic characteristics of these two types of
 AF-MPCs were examined by FACS analysis. Both types of
 AF-MPCs were negative for CD34, CD133, CD31, CD45, CD14 and
 HLA-DR. SS-AF-MPCs and RS-AF-MPCs were positive for MSC
 markers CD73, CD105 and CD166, adherent molecules CD29,
 CD44, CD49e and HLA-ABC (Fig. 2). The surface marker profiles
 were consistent with previously reported BM, AF and UCB hetero-
 geneous populations of MSCs [7, 9, 30, 31]. c-kit was expressed
 in similar very low/undetectable levels in both SS-AF-MPCs and
 RS-AF-MPCs populations, whereas CD146 was expressed in
 higher levels in RS-AF-MPCs compared to SS-AF-MPCs (Fig. 2).
 However, we observed that SS-AF-MPCs expressed high levels
 of CD90 [median fluorescent intensity (MFI): 58.33 ± 9.68]
 [Fig. 1B(vi)], whereas RS-AF-MPCs showed lower expression
 (MFI: 7.43 ± 6.53) [Fig. 1B(v)]. We further noticed that unselected
 cultures at passage 0 exhibited heterogeneous expression for
 CD90, low (MFI 1.6 ± 1.45) and high (MFI 53.4 ± 4.14), which
 may indicated the initial co-existence of the two different popula-
 tions [Fig. 1B(iv)]. More interestingly, the expression profile of SS-AF-
 MPCs and RS-AF-MPCs remained the same during culture, with

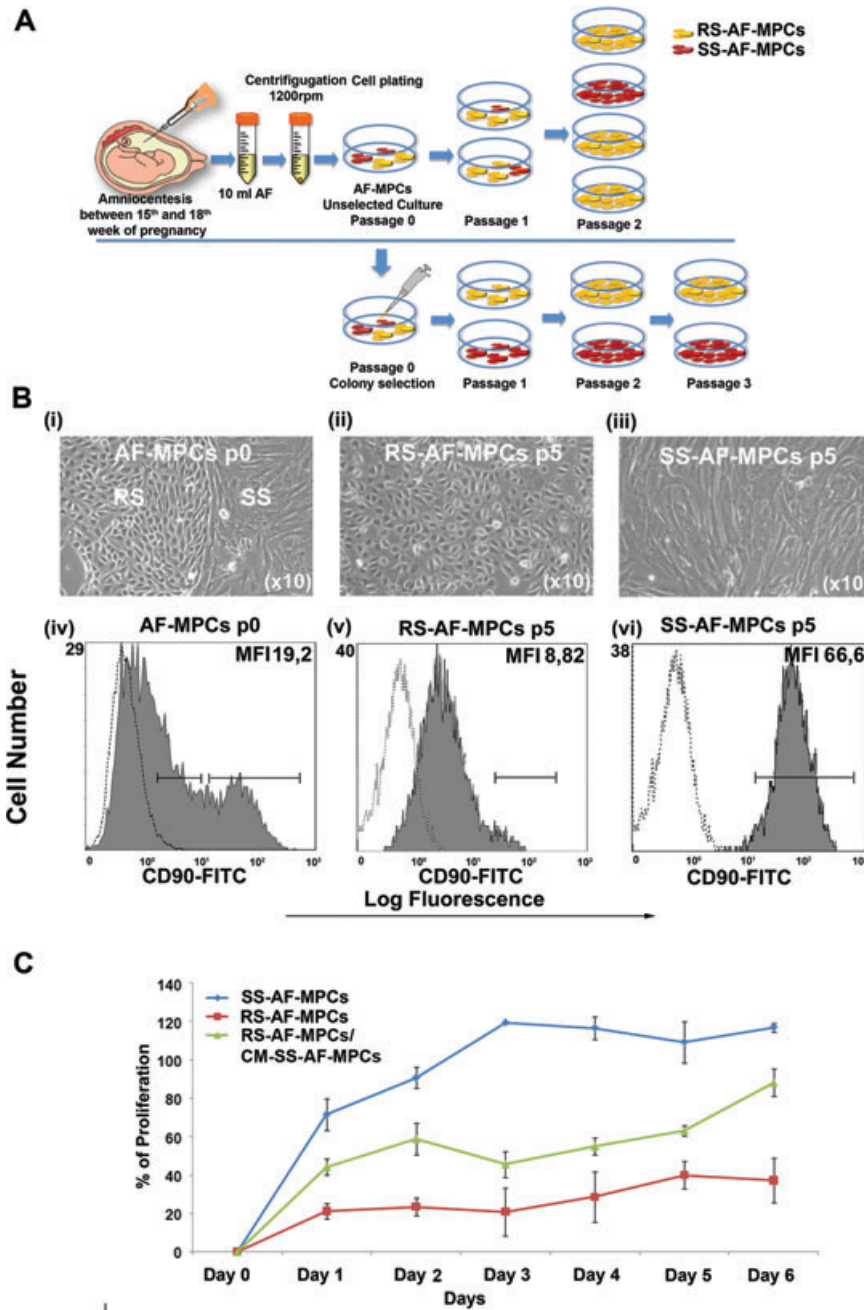


Fig. 1 Spindle- and round-shaped amniotic fluid mesenchymal progenitor cells. **(A)** Schematic representation of unselected AF-MPCs and mechanically selected colonies in an *in vitro* culture. **(B)** (i) AF-MPCs colonies morphology at p0, (ii) RS-AF-MPCs (p5) and (iii) SS-AF-MPCs (p5) morphology. Representative FACS histograms of (iv) AF-MPCs at p0, gated for CD90 expression (grey filled histograms), prior to analysis with the isotype-matched negative control (open histograms) at p0, (v) homogenous RS-AF-MPC at p5 and (vi) homogenous SS-AF-MPCs at p5 populations, respectively. **(C)** Comparative analysis of the percentage of proliferation of SS-AF-MPCs (blue line), RS-AF-MPCs (red line) and RS-AF-MPCs cultured in CM derived from SS-AF-MPCs (green line), during 6 days of culture by MTS assay. Values are mean \pm S.D. for three independent samples from each MPC population.

stable CD90 expression for each population, irrelevant to the culture passages and to the *in vitro* MPCs aging (data not shown). Another important observation was that SS-AF-MPCs exhibited high expression levels of CD44 adhesion molecule (MFI: 58.95 ± 8.91) compared to RS-AF-MPCs (MFI: 37.4 ± 10.46). We additionally examined the expression of MSC markers such as Vimentin and N-cadherin and also the epithelial marker E-cadherin. Both cell types were positive for Vimentin, whereas SS-AF-MPCs expressed E- and N-cadherin in a higher level (Fig. S1A).

CD90 expression altered according to the proliferation rate of SS-AF-MPCs

In an attempt to examine a potential relation of the enhanced expression levels of CD90 in SS-AF-MPCs with the high proliferation capacity they exhibit, we performed an *in vitro* proliferation assay at different temperature culture conditions. It is known from classical studies that when fibroblast-like cells are cultured at lower temperature (*i.e.* 33°C), they exhibit a lower proliferation rate [32].

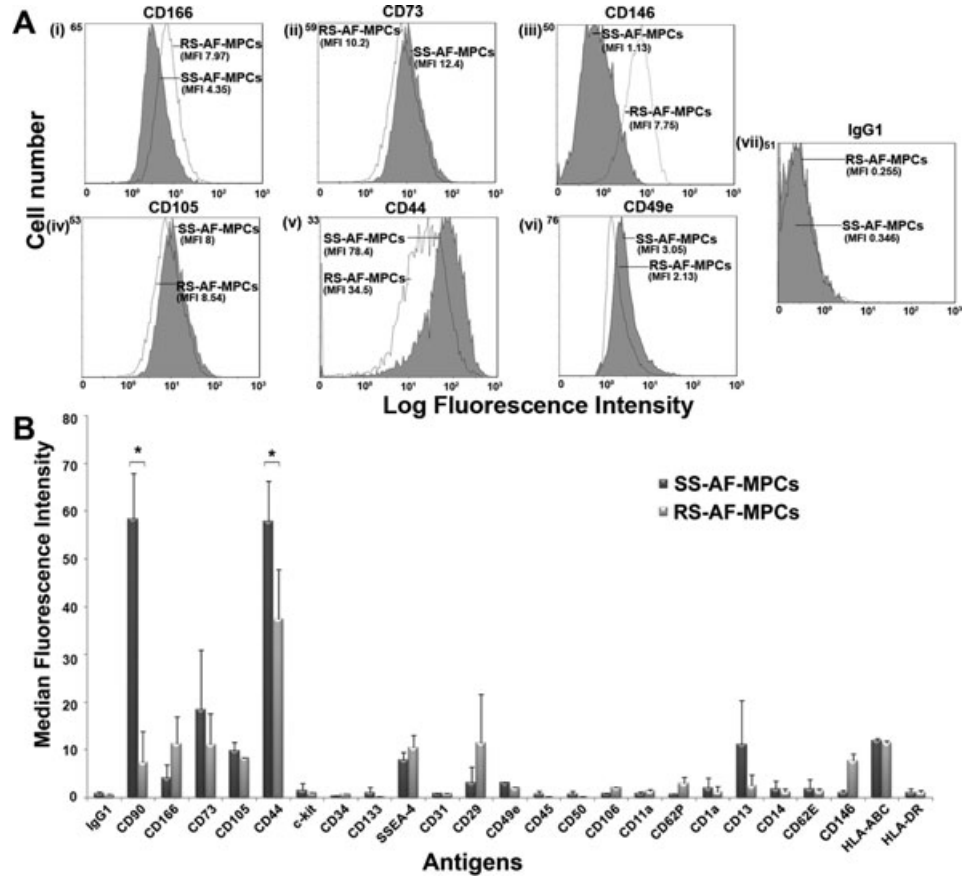


Fig. 2 Comparison of SS-AF-MPCs and RS-AF-MPCs expression patterns. (A) Representative FACS histograms of SS-AF-MPCs (grey filled histograms) and RS-AF-MPCs (opened histograms) gated for (i) CD166, (ii) CD73, (iii) CD146, (iv) CD105, (v) CD44 and (vi) CD49e markers, prior to analysis with (vii) the isotype-matched negative control. (B) SS- and RS-AF-MPCs analysed for different antigens expression by FACS analysis. The statistics were made on the mean MFI for each antigen. Isotype matched negative controls were used. Values are shown as mean \pm S.D. for three independent samples from each type. Statistical analysis was performed using the Student's t-test (* $P < 0.05$).

Therefore, we performed proliferation analysis assay for SS-AF-MPCs at 37°C (normal temperature conditions) and 33°C (low temperature conditions), respectively. The proliferation rate of SS-AF-MPCs dramatically decreased when cultured at 33°C for 6 days (Fig. 3A), compared to those cultured at 37°C for the same period. When we examined the expression of mesenchymal-related antigens at these time points, we observed that CD73 and CD105 levels were not altered when SS-AF-MPCs were cultured at 33°C (Fig. 3B). However, CD90 expression was decreased to 60.08% \pm 26.4 for day 1 [Fig. 3B(i)] and 62.09% \pm 3.65 for day 6 [Fig. 3B(ii)], respectively when the proliferation rate of SS-AF-MPCs was reduced because of temperature change. These results showed that CD90 antigen expression might be related to the growth rate of AF-MPCs and may in turn explain the difference of proliferation between the SS-AF-MPCs (CD90^{high}) and RS-AF-MPCs (CD90^{low}) subpopulations.

CD44 neutralizing antibody inhibited SS-AF-MPCs migration on fibronectin

Initially, a transwell migration assay was developed to determine the migration potential of SS-AF-MPCs and RS-AF-MPCs to

fibronectin and laminin. Interestingly, SS-AF-MPCs migrated approximately 7.59- and 7.79-fold faster towards fibronectin and laminin, respectively, than RS-AF-MPCs [Fig. 4A(i)] ($P < 0.001$, Student's t-test).

Taking under consideration that CD44 represented an adhesion molecule, binding on fibronectin and was expressed in SS-AF-MPCs in a higher level than RS-AF-MPCs according to FACS analysis (Fig. 2), we further investigated whether this elevated expression has a functional role in migration of SS-AF-MPCs. We therefore performed the same transwell migration assay after pre-incubating SS-AF-MPCs with a neutralizing antibody for CD44. Isotype non-binding antibody (IgG1) was used as a negative control. A significant decrease of 31.05% \pm 12.13 ($P < 0.05$, Student's t-test) in migration capacity of SS-AF-MPCs was observed in the presence of CD44 blocking antibody [Fig. 4A(ii)].

CD44 neutralizing antibody inhibited SS-AF-MPCs and RS-AF-MPCs adhesion on fibronectin and hyalouronic acid

SS-AF-MPCs and RS-AF-MPCs when tested in an *in vitro* adhesion assay on fibronectin and hyalouronic acid, exhibited similar

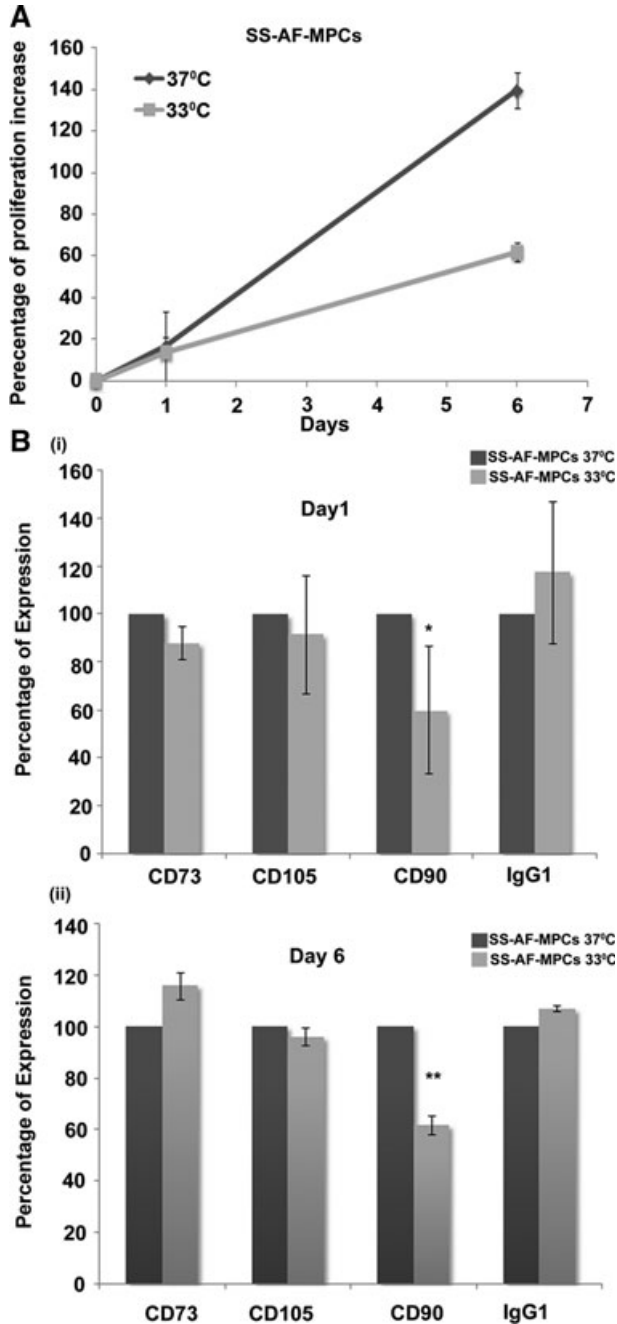


Fig. 3 CD90 expression alteration according to the proliferation rate of SS-AF-MPCs. (A) Comparative analysis of the percentage of proliferation increase of SS-AF-MPCs at 37°C (grey line) and 33°C (light grey line), respectively during 6 days of culture. (B) Comparison of the percentage of expression of CD73, CD105 and CD90 (i) at day 1 and (ii) day 6 of culture at 37°C (grey bars) and 33°C (light grey bars) by FACS analysis, respectively. MFI values were normalized for each marker against the level of expression determined at 37°C, which was set to 100%. Values are shown as mean \pm S.D. for three independent experiments. Statistical analysis was performed using the Student's t-test (* $P < 0.05$; ** $P < 0.001$).

adhesion capacities on both matrices [Fig. 4B(i-ii)-non-treated cells]. To analyse in detail the role of CD44 adhesion molecule, which binds to fibronectin and also to hyalouronic acid [33], we examined whether CD44 controlled not only the high migration capacity of SS-AF-MPCs as shown before, but also the adhesion of both populations to the respective binding matrices. Blocking of CD44 resulted in lower adhesion of both SS-AF-MPCs and RS-AF-MPCs on fibronectin at approximately 48% ($P < 0.05$, Student's t-test) and 31.3% ($P < 0.05$, Student's t-test), respectively [Fig. 4B(i)]. However, only SS-AF-MPCs exhibited impaired adhesion on hyaluronic acid in the presence of CD44 blocking antibody, (37.9%, $P < 0.05$, Student's t-test). There was no significant effect on cell adhesion using the IgG1 antibody compared to the non treated cells in both matrices [Fig. 4B(i) and (ii)].

CD49e modulated SS-AF-MPCs and RS-AF-MPCs adhesion properties

Both cell types express the CD49e molecule which binds to fibronectin, as determined by FACS analysis (Fig. 2). For examining the role of CD49e on fibronectin-mediated adhesion, we analysed SS-AF-MPCs and RS-AF-MPCs in an *in vitro* adhesion assay. We observed an approximately 87% ($P < 0.001$, Student's t-test) and 68.9% ($P < 0.001$, Student's t-test) reduction on adhesion ability of SS-AF-MPCs and RS-AF-MPCs, respectively, in the presence of CD49e blocking antibody, compared to non-treated cells [Fig. 4B(i)].

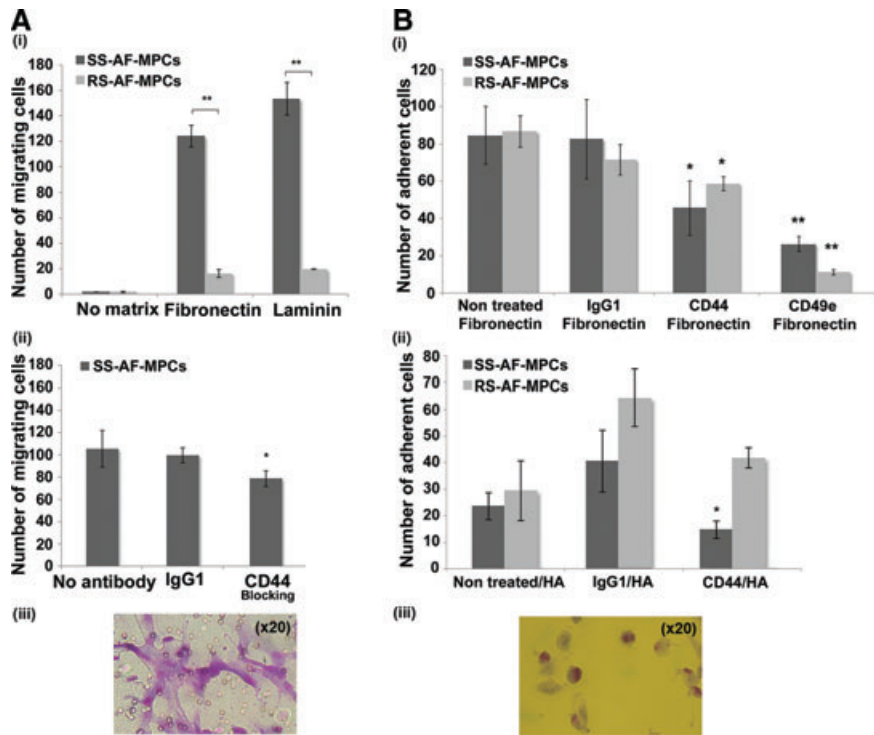
SS-AF-MPCs expressed oct-4, nanog and sox-2 in higher levels than RS-AF-MPCs

We next examined the expression of the oct-4, sox-2 and nanog transcripts, key markers in embryonic stem cells and responsible for the maintenance of pluripotency of mammalian stem cells *in vivo* and *in vitro*, in SS-AF-MPCs and RS-AF-MPCs of passages 5–7 [34, 35]. Oct-4, nanog and sox-2 exhibited a 2.6 ± 1.38 , 10.41 ± 3.38 and 15.18 ± 3.92 fold higher expression respectively, in SS-AF-MPCs compared to RS-AF-MPCs, by RT-PCR analysis after normalization to GAPDH endogenous control (Fig. 5A). However, both AF-MPCs types expressed Oct-4 and Sox-2 at protein level, as demonstrated by nuclear immunofluorescent staining (Fig. 5B). In all samples examined, the majority of the cells were positive for Oct-4 and Sox-2.

SS-AF-MPCs exhibited different differentiation properties compared to RS-AF-MPCs

SS-AF-MPCs and RS-AF-MPCs were cultured under appropriate conditions that induce adipocytes, osteocytes, chondrocytes (mesoderm-derived) and hepatocytes (endoderm-derived), to evaluate their *in vitro* differentiation properties (Fig. 6). Results

Fig. 4 Migration and adhesion properties of SS-AF-MPCs and RS-AF-MPCs. **(A)** (i) SS-AF-MPCs showed higher motility ($P < 0.001$) on fibronectin and laminin, respectively compared to RS-AF-MPCs. (ii) Number of migrated SS-AF-MPCs to fibronectin in presence of CD44 neutralizing antibody or isotype matched non-specific antibody IgG1. (iii) Representative image (20 \times) of migrated SS-AF-MPCs fixed and stained using the Ral staining kit on the transwell membrane. **(B)** (i) Number of adherent SS-AF-MPCs and RS-AF-MPCs to fibronectin, treated with CD44, CD49e neutralizing antibodies or isotype matched non-specific antibody IgG1 in comparison to non treated SS-AF-MPCs and RS-AF-MPCs, respectively. (ii) Number of adherent SS-AF-MPCs and RS-AF-MPCs to hyalouronic acid, treated with CD44 neutralizing antibody or isotype matched non-specific antibody IgG1 in comparison to non treated SS-AF-MPCs and RS-AF-MPCs, respectively. (iii) Representative image (20 \times) of adherent cells fixed and stained using the Ral staining kit on the plastic vessel. Values are shown as mean \pm S.D. for three independent experiments. Statistical analysis was carried out using the Student's t-test ($*P < 0.05$; $**P < 0.001$).



showed that RS-AF-MPCs exhibited an enhanced adipogenic (1.6 ± 0.4 fold times increase) [Fig. 6A (iii)] and reduced osteogenic (4.6 ± 1.0 fold times decrease) [Fig. 6B(iii)] differentiating capacity compared to SS-AF-MPCs, as determined by oil red O and alkaline phosphatase staining and quantified by Image J analysis, respectively. Interestingly enough, under chondrogenic and hepatogenic inducing conditions, RS-AF-MPCs failed to form chondrocytes and hepatocyte-like cells respectively, in contrast to SS-AF-MPCs (Fig. 6C and D). Chondrogenesis, was determined by Alcian Blue staining of pellets (Fig. 6C). Hepatogenesis was verified by PAS staining, where hepatocyte-like cells derived from SS-AF-MPCs exhibited a 73.07 ± 5.2 fold times increase in PAS staining compared to SS-AF-MPCs [Fig. 6D(ii)], urea production ($46.5 \text{ mg/dl} \pm 10.2$) compared to SS-AF-MPCs ($9.6 \text{ mg/dl} \pm 6.4$) [Fig. 6D(iii)] and high albumin expression level (12.5 ± 2.4 fold expression difference compared to SS-AF-MPCs) [Fig. 6D(iv)].

Differentially expressed proteins identified in SS-AF-MPCs and RS-AF-MPCs

To further characterize the AF-MPCs subpopulations, proteins differentially expressed in SS-AF-MPCs and RS-AF-MPCs were identified by 2D-gel electrophoresis and MS. Total protein extracts of three different SS-AF-MPCs and RS-AF-MPCs preparations were analysed in duplicate by 2DE in 4–7 pH gradient strips. The derived pattern of resolved protein spots for each cell type was highly consistent and a comparison of the expression levels of the respective

proteins was established using PDQuest 8 software (Fig. 7A and B). Protein spots that were found to be more than 1.5 times fold differentially expressed and at statistically significant levels ($P < 0.05$ according to Student's t-test and/or Mann-Whitney test) in the two subpopulations, are described in detail in Tables S1 and S2. Specifically, proteins up-regulated in SS-AF-MPCs compared to RS-AF-MPCs included reticulocalbin-3 precursor, collagen $\alpha 1$ (I) chain precursor, FK506-binding protein 9 precursor, Rho GDP-dissociation inhibitor 1, chloride intracellular channel protein 4, tryptophanyl-tRNA synthetase and heat shock proteins 1 70 kDa and $\beta 1$. On the other hand, proteins such as peroxiredoxin-2, 60 kDa heat shock protein, glutathione S-transferase P and annexin A4, were up-regulated in RS-AF-MPCs. Proteins identified only in RS-AF-MPCs included cytoke- ratin 8, 18 and 19, cathepsin B, coactosin-like protein and integrin α -V protein (Table S3).

Among the proteins overexpressed in SS-AF-MPCs, collagen $\alpha 1$ (I), a major extracellular matrix molecule responsible for directing the fate of MSCs into osteogenic lineages [36], was the most abundant. This finding may in turn explain the enhanced differentiation potential of SS-AF-MPCs to osteoblastic lineages, as observed in our *in vitro* studies. Proteins facilitating cell migration, such as Rho GDP-dissociation inhibitor 1 [37] and tryptophanyl-tRNA synthetase [38] are expressed in higher levels in SS-AF-MPCs, reflecting their enhanced motility *in vitro*. On the other hand, the high proliferation rate of SS-AF-MPCs might be explained by the presence of proteins such as chloride intracellular channel protein 4, found to promote endothelial cell proliferation and cell survival [39]. Similarly, heat shock protein $\beta 1$, exhibiting

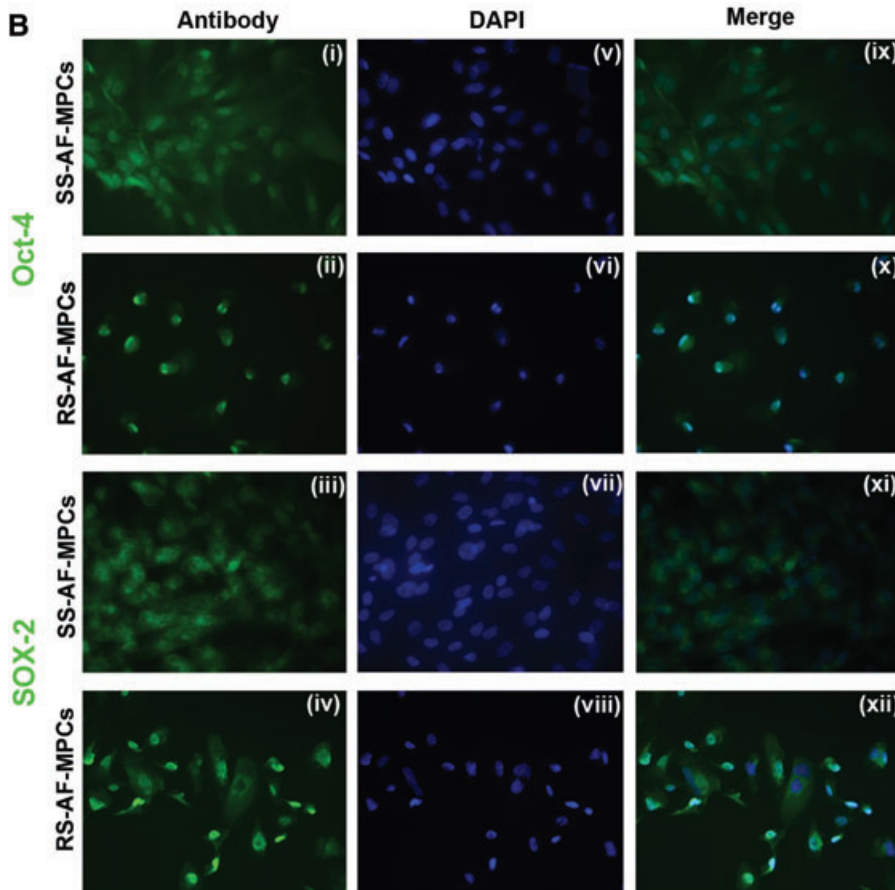
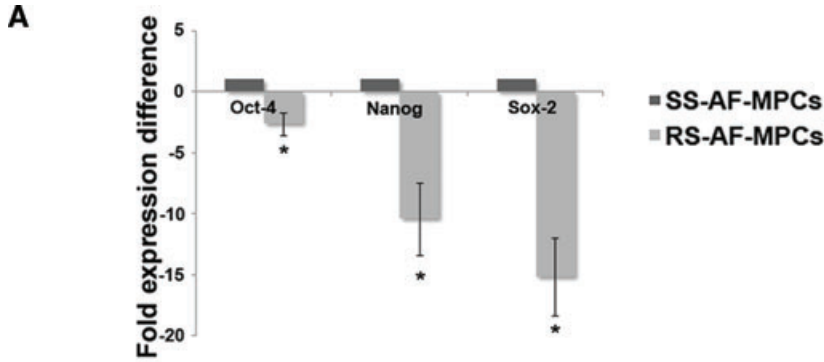


Fig. 5 Embryonic stem cell marker expression and differentiation potential of SS-AF-MPCs and RS-AF-MPCs. **(A)** Comparative analysis for the expression of oct-4, nanog and sox-2 in three SS-AF-MPCs and RS-AF-MPCs samples, respectively, analysed by RT-PCR. Results were first normalized to human GAPDH positive control and then to SS-AF-MPCs expression levels for each marker, respectively. Statistical analysis was performed using the Student's t-test. **(B)** Immunofluorescent nuclear staining for (i–ii) Oct-4, (iii–iv) Sox-2 and (v–viii) DAPI (ix–xii) of SS-AF-MPCs and RS- AF-MPCs. Merge of DAPI staining and antibody staining. Original magnifications, 40 \times .

an anti-apoptotic role by former studies, was also highly expressed in SS-AF-MPCs [40, 41].

Confirmation of the differentially expressed proteins CK18, CK19, cathepsin B and collagen α 1 (I)

RS-AF-MPCs exhibited a more complicated protein profile compared to SS-AF-MPCs, including 10 uniquely expressed proteins,

supporting the indication of an earlier developmental stage of the SS population. To verify the 2DE results and confirm the differential expression, cytokeratin 18, cytokeratin 19 and cathepsin B were further analysed by Western blotting. It was demonstrated that bands of 40, 54 and 28 kDa, corresponding to CK19, CK18 and cathepsin B, respectively were only detected in RS-AF-MPCs (Fig. 7C). Furthermore, collagen α 1 (I) was expressed in higher levels in SS-AF-MPCs than in RS-AF-MPCs again in agreement to the 2DE results (Fig. 7D).

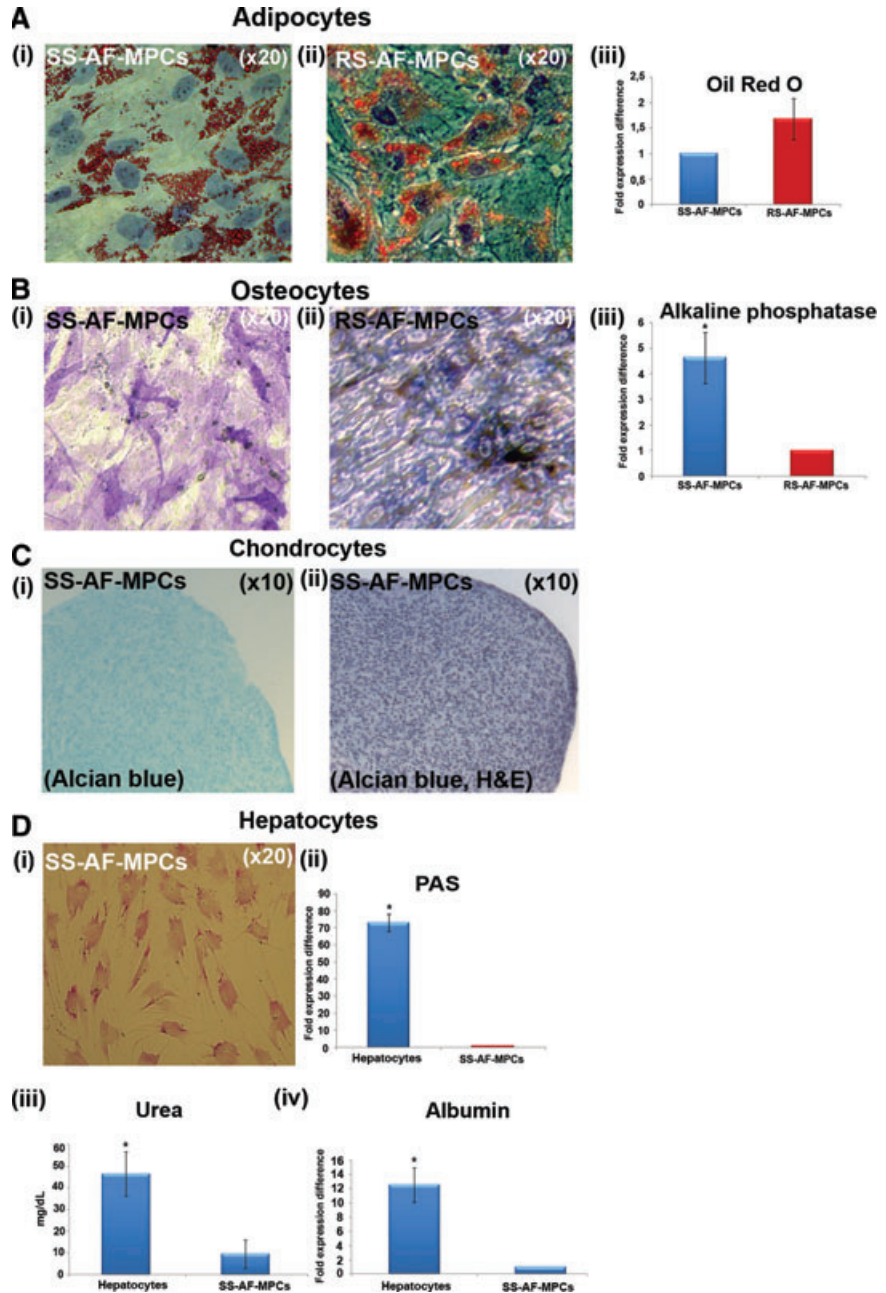


Fig. 6 (A) Oil Red O staining for adipocyte differentiation for (i) SS-AF-MPCs and (ii) RS-AF-MPCs respectively, followed by (iii) quantitation analysis. (B) Alkaline phosphatase staining for osteocyte differentiation for (i) SS-AF-MPCs and (ii) RS-AF-MPCs respectively, followed by (iii) quantitation analysis. (C) (i) Alcian Blue and (ii) haematoxylin and eosin staining of SS-AF-MPCs, cultured under chondrogenic inducing conditions in pellet mass cultures. (D) PAS staining for hepatocyte differentiation for (i) SS-AF-MPCs induced to hepatocytes. (ii) Quantitation analysis for PAS staining, (iii) determination of the secreted Urea and (iv) albumin expression were shown. Quantitation of the respective differentiation assays was performed by using the Image J analysis software on 10 fields per image. For each sample, four images were taken. For adipogenic and osteogenic differentiation values were normalized in each case against the AF-MPC type with the lower differentiation capacity, which was set to 1, whereas for hepatogenic differentiation, values were normalized in each case against non-induced to differentiation SS-AF-MPCs. Values are mean \pm S.D. from three samples from each type. Statistical analysis was performed using the Student's t-test, $P < 0.05$.

Lentivirus-mediated gene transfer of AF-MPCs

Taking under consideration the successful, easy and rapid expansion of SS-AF-MPCs in culture, as well as the high passage they reach compared to RS-AF-MPCs, we further investigated whether they can be efficiently transduced with lentiviral vectors for further potential use in *in vivo* therapeutic applications. For this reason, SS-AF-MPCs were transduced with GFP third generation lentivirus [28]. The experimental procedure was focused on three parameters:

first, the efficiency of same passage SS-AF-MPCs transduction by a virus dose-dependent manner; secondly, the efficiency of GFP expression in different passages and thirdly, the long-term maintenance of GFP expression in culture. An increase in fluorescence of GFP positive SS-AF-MPCs was shown, when the cells, at passage 16, were transduced with rising doses of virus, MOI from 10 to 100, as determined by FACS analysis four days post-transduction (Fig. 8A and B). The transduction with an MOI of 60 led to a 98–100% efficiency of infection. A very small percentage of cells

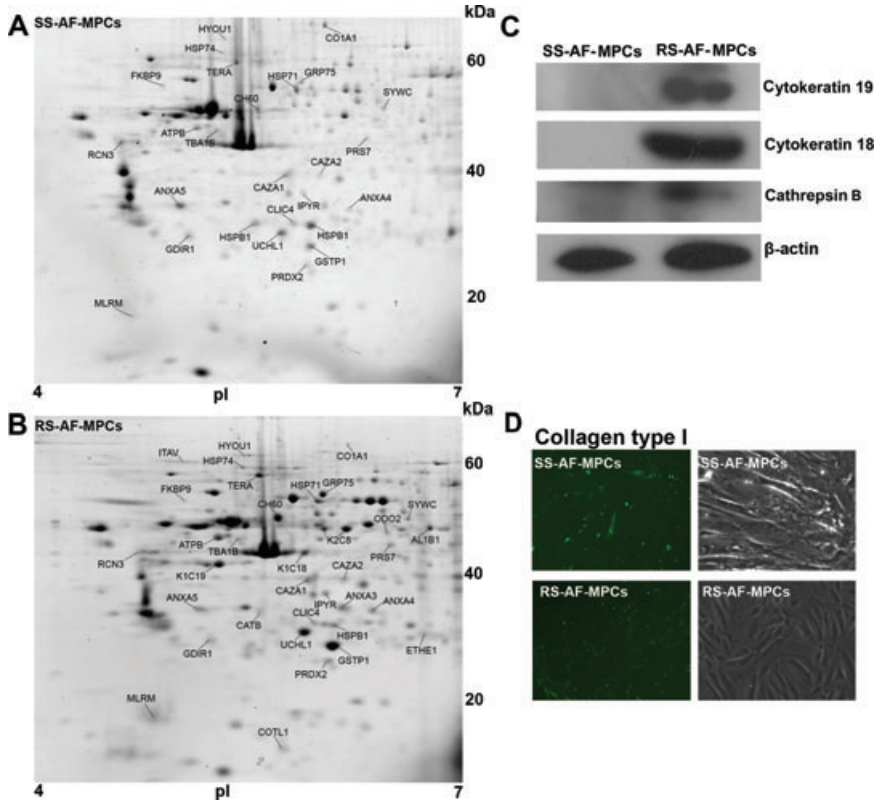


Fig. 7 Two-dimensional gel electrophoretic analysis of AF-MPCs. **(A)** Representative 2D-gel electrophoresis image of proteins extracted from SS-AF-MPCs and **(B)** RS-AF-MPCs. The differentially and unique expressed protein spots in each population are indicated with their abbreviated names and listed in Tables S1–3, respectively. **(C)** Confirmation of the cytoskeleton 19 and 18 and cathepsin B expression by Western blot analysis with the respective antibodies in cell extracts from SS-AF-MPCs (lanes 1) and RS-AF-MPCs (lanes 2). Protein bands of 40, 54 and 38 kDa corresponding to cytoskeleton 19 and 18 and cathepsin B heavy chain were detected. Immunoblotting for b-actin has been conducted to ensure the comparable loading of proteins in each lane. **(D)** Confirmation of the higher expression of collagen α 1 (I) protein in SS-AF-MPCs compared to RS-AF-MPCs by immunofluorescent staining.

were positive for Annexin V staining only in virus dosages higher than MOI 60 (Fig. 8C). The expression levels of mesenchymal markers (CD90, CD105 and CD73) in SS-AF-MPCs remained the same after transduction (data not shown).

Furthermore, we performed a different passage expression analysis using SS-AF-MPCs of passages 10–40, transduced with an MOI of 60 (Fig. 8D). Interestingly enough, the passage did not affect the expression of GFP. In addition, the expression of GFP was maintained at the same high levels 4 weeks and 9 months post-transduction (Fig. 8E). By series of freezing and thawing cycles we observed that the expression of GFP remained constant. In addition, GFP-SS-AF-MPCs (MOI 60) exhibited high proliferation capacity (Fig. S1B) and were still capable to differentiate into adipogenic, osteogenic and hepatogenic lineages *in vitro*, as shown in Figure S1C.

In vivo engraftment of left ventricular transduced SS-AF-MPCs

The high proliferation rate of SS-AF-MPCs and the fact that they exhibit normal karyotype at high passages, allowed the use of these cells *in vivo*. RS-AF-MPCs were not efficient in number to be used in *in vivo* experiments.

Thus, we further investigated the *in vivo* survival, migration and engraftment of GFP-SS-AF-MPCs in non-damaged immunosuppressed mice. GFP-SS-AF-MPCs were *i.v.* injected into

NOD/SCID mice and then analyses involving FACS, immunohistochemistry and RT-PCR for the WPRE gene were used, to follow the distribution of donor cells 4 and 10 days post-injection at different tissues (Fig. 9). For immunohistochemistry anti-human CD90 antibody, which does not cross-react with mouse MSCs and tissue (Fig. S1D) was also used. We were able to detect GFP-SS-AF-MPCs at liver, spleen, lung, gut and kidney at these time points. However, similarly to previous studies with MSCs [42], we could not detect SS-AF-MPCs in the BM as confirmed by FACS analysis (data not shown). No tumours were detected even after 3 months post-SS-AF-MPCs transplantation in NOD/SCID animals, which may indicate non-tumorigenic properties of these cells.

Furthermore, we tested the viability of SS-AF-MPCs supported by matrigel and transplanted subcutaneously into the tail bases of NOD/SCID mice. It was demonstrated by fluorescent microscopy and FACS analyses that matrigel succeeded in supporting SS-AF-MPCs engraftment for at least 10 days without losing the GFP expression (Fig. 9D). This finding proved that extracellular components provided by matrigel were needed to improve SS-AF-MPCs survival and engraftment *in vivo*.

Discussion

Recent interest is focused on AF as a valuable source of MPCs [7–9, 11, 12]. Culture of AF-MPCs, as previously reported

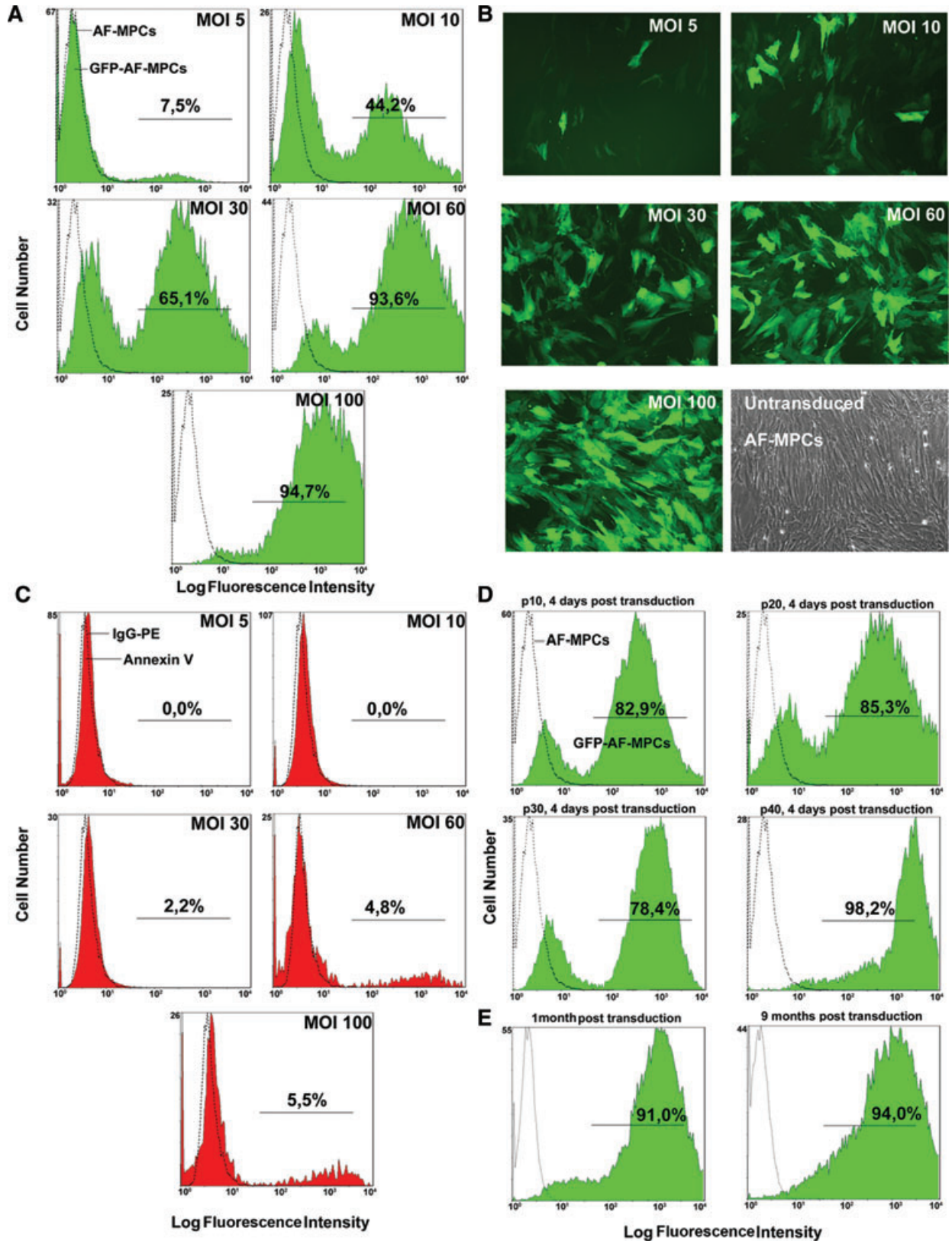




Fig. 8 Transduction of SS-AF-MPCs with the pCCLsin.PPT.hPGK.GFP lentiviral system. **(A)** Flow cytometric analysis, **(B)** microscopic evaluation (20×) and **(C)** evaluation of apoptosis by Annexin V staining by FACS analysis of SS-AF-MPCs at MOI 5–100 and passage 16 **(D)** GFP efficiency of SS-AF-MPCs determined 4 days post-transduction at passages 10, 20, 30 and 40 by FACS analysis of MOI 60. **(E)** Stability of GFP expression 1 and 9 months post-transduction by FACS analysis at MOI 60.

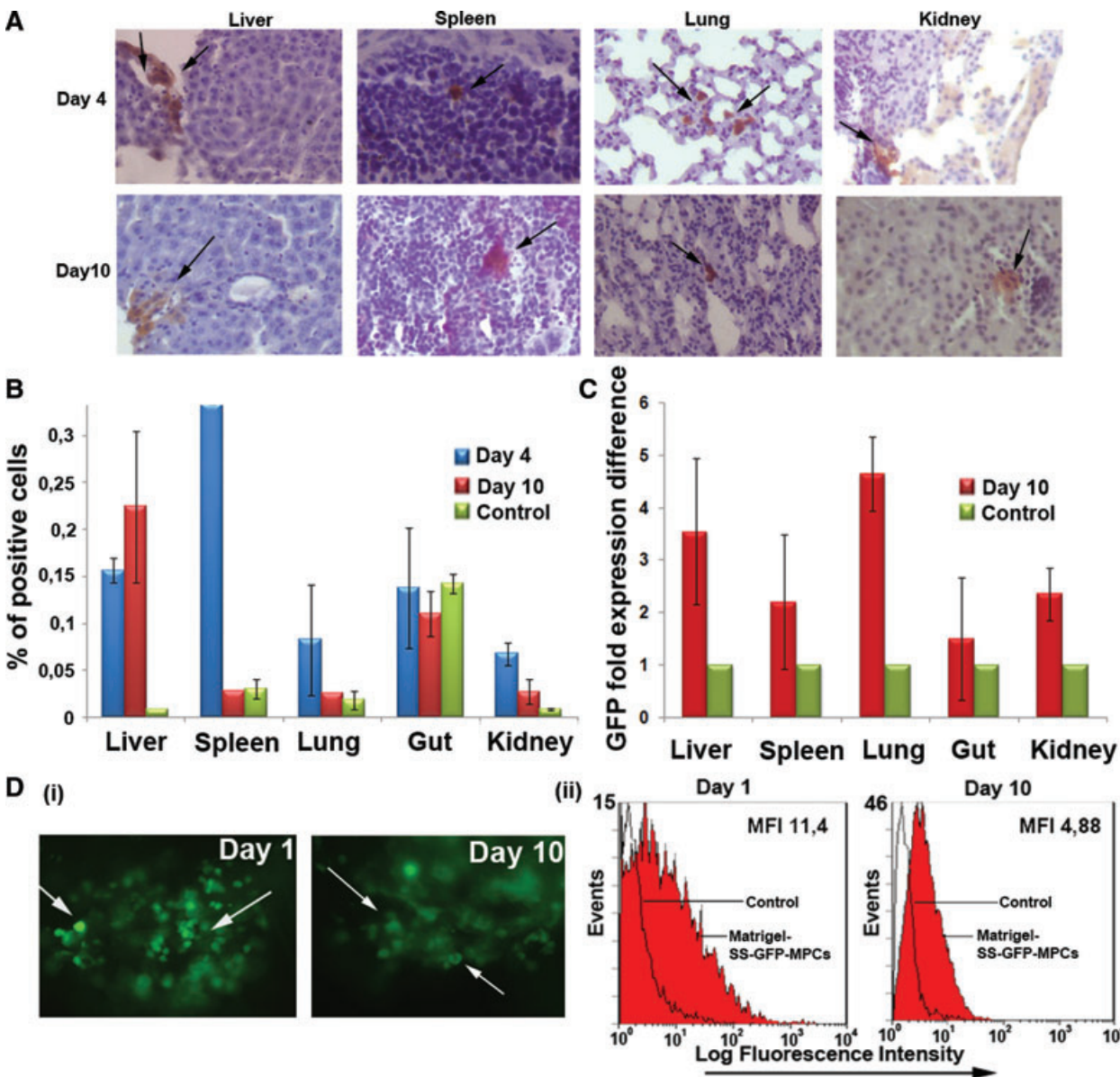


Fig. 9 *In vivo* engraftment of GFP transduced SS-AF-MPCs. **(A)** GFP-SS-AF-MPCs were trapped in different tissues as evaluated by immunohistochemistry, 4 and 10 days after transplantation. GFP-SS-AF-MPCs were found in liver, spleen, lung and kidney at low frequency. Immunohistochemistry was performed by using anti-GFP antibody. Quantitation of GFP cells was determined by **(B)** FACS analysis and **(C)** RT-PCR in the respective tissues 4 or 10 days post-injection where percentage of positive cells and GFP fold expression difference is presented, respectively. As negative controls, non-injected mice were used. Values are shown as mean \pm S.D. for four mice in each group. **(D)** (i) Representative post-mortem fluorescent microscopy image of SS-AF-MPCs within the matrigel revealed a robust engraftment. (ii) FACS analysis of the disassociated matrigel area in presence of GFP transduced SS-AF-MPCs (red filled histogram) or GFP transduced SS-AF-MPCs in PBS (open histogram) 1 and 10 days post-transplantation.

1 from our group and others, yielded in an adherent heterogenic
2 cell population with diverse morphology [7, 8, 11]. The
3 expression analysis, determined by FACS analysis, documented
4 the presence of CD44, CD90, CD73 and CD105 and the
5 absence of haematopoietic-related CD-antigens [7, 8, 11],
6 resembling to the phenotype of MSC-like cells according to
7 standard criteria [43].

8 In our previous studies, we successfully isolated and expanded
9 karyotypically normal MSCs from 80 samples of second trimester
10 AF and performed a systematic phenotypic, molecular and pro-
11 teomic analysis [7]. The main characteristic of the AF-MSCs was
12 the high number of isolated cells and their rapid expansion *in vitro*
13 compared to BM-MSCs. More importantly, these cells when
14 exposed to appropriate differentiation media *in vitro*, showed a
15 multilineage differentiation potential and ability to overcome the
16 mesodermal commitment, by differentiating into cell types derived
17 from all three germ layers [7].

18 In this study, we focused on the detailed investigation of differ-
19 ent populations of AF-MPCs. We initially observed that unselected
20 cultures contained, at passage 0–1, two morphologically distinct
21 populations, SS-AF-MPCs and RS-AF-MPCs, whereas at passage
22 2–3 the RS cells were more abundant in 94% of the samples. The
23 rest 6% of the samples represented a SS cell population that can
24 be expanded up to 40–50 passages to date. In an attempt to char-
25 acterize better these two subpopulations, we randomly chose six
26 AF samples at passage 0, where a mixture of both cell types
27 existed at almost equal frequency, then we mechanically isolated
28 SS or RS colonies and successfully expanded them under the
29 same culture conditions.

30 Although cultures of MPCs have been studied extensively the
31 last decades, standard criteria for isolating and expanding these
32 cells have not been developed [4]. Several protocols have been
33 established for isolating human MPCs from different sources by
34 using staining with different antibodies [3, 4, 6–9, 11, 13, 16, 44].
35 Other groups have reported that the existence of morphologically
36 distinct MPC subpopulations is related to the plating density of the
37 sample [1, 4, 44]. However, in this study, we observed the pres-
38 ence of both RS-AF-MPCs and SS-AF-MPCs at passage 0 despite
39 the initial plating density.

40 A detailed examination of the phenotype of SS-AF-MPCs and
41 RS-AF-MPCs showed that SS-AF-MPCs expressed higher levels
42 of CD90. However, the unselected AF-MPCs cultures at passage
43 0–1 exhibited a mixed expression of CD90 revealing the initial co-
44 existence of both SS and RS cells. This observation was consistent
45 with older reports on murine lung fibroblasts and UCB-MPCs, in
46 which two morphologically different populations were identified,
47 one SS and CD90 positive, and the other one rounded and CD90
48 negative. Furthermore, it has been shown that CD90 expression
49 affected the morphology, proliferation and differentiation of fibro-
50 blasts [45]. RS-AF-MPCs shared similar morphological characteris-
51 tics to human amniotic membrane epithelial cells (hAEC), which
52 express stem cell markers and have the ability to differentiate to
53 multiple cell lineages [46]. However, hAEC differentiate success-
54 fully to hepatocytes and exhibit increased expression of CD90 after
55

second passage in culture, in contrast to RS-AF-MPCs [46].
In addition, SS-AF-MPCs shared the rapid expansion and the
multi-lineage differentiation potential of human umbilical cord
perivascular cells (HUCPVCs), whereas they exhibited low expres-
sion of CD146, a marker characteristic for circulating endothelial
cells [47].

We further decided to study in more detail the molecular
identity and specific properties of the SS-AF-MPC population.
We observed that CD90 expression was decreased when the
proliferation rate of SS-AF-MPCs was reduced because of tem-
perature change. These results showed that CD90 antigen
expression could be related to the growth rate of AF-MPCs and
may in turn explain the difference of proliferation between
SS-AF-MPCs (CD90^{high}) and RS-AF-MPCs (CD90^{low}) subpopula-
tions. However, temperature change may also affect other
biological parameters and for this reason further investigation is
needed to explore the signaling pathway that is possibly related
to this observation.

Furthermore, despite the similar stem cell features of SS-AF-
MPCs and RS-AF-MPCs, the first ones possess a significantly
higher migration capacity, compared to RS-AF-MPCs. This find-
ing provided a basis for a more extended investigation of the
migration mechanism of SS-AF-MPCs, involving a variety of
adhesion molecules such as VLA-4, VLA-5, CD29 or CD44 [48].
For example, blocking studies by using CD44 neutralizing
antibody showed impaired migration properties of SS-AF-MPCs
on fibronectin.

SS-AF-MPCs and RS-AF-MPCs expressed the pluripotency
markers Oct-4 and Sox-2, indicating a possible primitive pheno-
type and stem cell potential of these cells. Similar findings by
Guillot *et al.* [7, 49] for MSCs isolated from first-trimester foetal
blood, liver and BM confirmed that MSCs derived from neonatal
and mid-gestational foetal tissues also expressed pluripotency
markers, such as Oct-4 and Sox-2. In parallel, comparative
studies by Greco *et al.* [50], showed that MSCs and embryonic
stem cells (ESCs) shared similar expression of the embryonic
transcription factors Oct-4, Sox-2 and Nanog, both at RNA and
protein levels.

In addition, by the use of proteomic analysis, 25 proteins were
found differentially expressed among the two subpopulations,
which might explain the discrepancy in their proliferation, migration
and differentiation properties. The differentially expressed proteins
between SS-AF-MPCs and RS-AF-MPCs did not include any of the
proteins highly expressed in MSCs from various sources such as
Vimentin, Galectin, Gelsolin and Prohibitin [7, 47].

Our data indicated for the first time that SS-AF-MPCs are
highly susceptible to lentiviral transduction, with no silencing
effects over the multiple culture passaging. Moreover, GFP
expression was retained through *in vitro* differentiation of SS-
AF-MPCs, indicating the prospective utilization of this type of
MPCs in gene therapy applications. Previous studies showed
that human AF stem cells, seeded in a scaffold and exposed to
osteogenic-inducing medium, can form bone *in vivo* [9].
However, up to date there have been no conclusive reports

demonstrating the *in vivo* contribution of AF-MPCs upon intravenous delivery without tissue damage of the recipient animals. Here, we report that the SS-AF-MPCs can home into several tissues, at low frequency, after systemic infusion into recipient animals and can be tracked in lung, liver or spleen after several days. More importantly, we have demonstrated that matrigel supported SS-AF-MPCs engraftment is superior to cells alone. This can be explained by the specific properties of the matrigel basement, providing growth factors and structural support to the cells [6, 45, 51–53].

Therefore, we succeeded in mechanically isolating and expanding two mesenchymal origin populations of AF (SS-AF-MPCs and RS-AF-MPCs) that co-exist at early passage. SS-AF-MPCs are characterized by higher CD90 antigen expression than RS-AF-MPCs. The ease with which SS-AF-MPCs can be expanded in culture represents a marked contrast to the difficulties that have been encountered in expanding MPCs derived from sources such as BM, AF or UCB [6, 7, 9, 11, 31]. The specific properties of SS-AF-MPCs related to migration ability, differentiation capacity, lentiviral transduction efficiency and long-term survival *in vivo* will be of great importance in their use for cell and gene therapy. Under the conditions developed in this study, from a 10 ml AF sample we can mechanically isolate SS-AF-MPCs colonies that can generate millions of cells in culture in a short period of time, enough for future clinical applications.

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Conflict of interest

The authors confirm that there are no conflicts of interest.

Supporting information

Additional Supporting Information may be found in the online version of this article:

Fig. S1 (A) Immunofluorescent nuclear staining for E-cadherin, N-cadherin and Vimentin proteins merged with DAPI staining of SS- and RS- AF-MPCs. Original magnifications, 40 \times . **(B)** Comparative analysis of the percentage of proliferation of SS-AF-MPCs (blue line) and GFP-SS-AF-MPCs (red line) during 7 days of culture. **(C)** GFP transduced SS-AF-MPCs differentiation to adipocyte, osteocytes and hepatocytes. **(D)** Representative FACS histograms of SS-AF-MPCs (black filled histograms) and mouse BM-MSCs (grey filled histograms) gated for CD90, prior to analysis with the isotype-matched negative controls (blue and red open histograms), respectively. **(E)** Normal karyotype of (i) SS-AF-MPCs and (ii) RS-AF-MPCs from two representative samples derived from male [46XY(20)] and female embryo [46XX(20)], respectively at passage 5. (iii) Normal karyotype of SS-AF-MPCs from 1 representative sample derived from female embryo [46XX(20)], at passage 32. Forty metaphase spreads were fully analysed and karyotyped in each case.

Table S1 Proteins up-regulated in SS-AF-MPCs

Table S2 Proteins up-regulated in RS-AF-MPCs

Table S3 Proteins expressed in RS-AF-MPCs only

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